Naval Research Laboratory

Washington, DC 20375-5320



NRL/MR/6110--01-8599

Fiber Optic Raman Spectroscopy for Detection of Methane Hydrates and Related Species

SEAN J. HART ROBERT A. LAMONTAGNE

Chemical Dynamics and Diagnostics Branch Chemistry Division

December 19, 2001

20020110 148

Approved for public release; distribution is unlimited.

REPORT DOCUMENTATION PAGE

Form Approved OMB No. 0704-0188

Public reporting burden for this collection of information is estimated to average 1 hour per response, it

,	ns for reducing this burden, to Washing 202-4302, and to the Office of Manage	action of information. Send comments rega gton Headquarters Services, Directorate fo ement and Budget, Paperwork Reduction F	eviewing instructions, searching existing data sources, arding this burden estimate or any other aspect of this or Information Operations and Reports, 1215 Jefferson Project (0704-0188). Weakington		
AGENCY USE ONLY (Leave Blank)	2. REPORT DATE		REPORT TYPE AND DATES COVERED		
	December 19, 2001	Interim			
4. TITLE AND SUBTITLE			1-		
Fiber Optic Raman Spectrosco	opy for Detection of Methane	Hydrates and Related Species	5. FUNDING NUMBERS		
6. AUTHOR(S)					
Sean J. Hart and Robert A. La					
7. PERFORMING ORGANIZATION NAM	ME(S) AND ADDRESS(ES)		8. PERFORMING ORGANIZATION		
Naval Research Laboratory	Naval Research Laboratory				
4555 Overlook Avenue, SW Washington, DC 20375-5320	NRL/MR/611001-8599				
9. SPONSORING/MONITORING AGENC	CY NAME(S) AND ADDRESS(ES)				
Office of Naval Research	المارات	1	10. SPONSORING/MONITORING AGENCY REPORT NUMBER		
800 North Quincy Street Arlington, VA 22217					
Armigion, VA 22211					
11. SUPPLEMENTARY NOTES					
-					
12a. DISTRIBUTION/AVAILABILITY STAT	FEMENT		12h DISTRIPLETION COOP		
Annroyed for public release, di	e a en carrolla de la carrolla de l En altre en carrolla de la carrolla		12b. DISTRIBUTION CODE		
Approved for public release; di	stribution is unlimited.				
13. ABSTRACT (Maximum 200 words)					
(maximum 200 Worus)					
of sample interrogated by the ex in the capillary which is transmi this system for methane hydrate methane in water at atmospher hydrate at great depths (1000's hydrate may be present. The rela- mented. The benefit of using a cap	ccitation light. A Nd: YAG lase itted by the collection fiber to e detection is evaluated througic pressures. The eventual approximate the collection is evaluated through the formula of the collection of the co	er operating at 532 nm is used to go a notch filter holder and linear are the use of organic surrogate mapplication is the oceanic detection and the solubility of methan from the methane and other organic complication is the oceanic detection of the solubility of methans and other organic complications of the solubility of methans and other organic complications.	frapidly collecting Raman spectra of guide designed to increase the volume generate Raman scatter at the sample generate Raman scatter at the sample generate. The feasibility of using toolecules, due to the low solubility of on of methane gas, liquid, and solid gane increases significantly and solid pounds relative to nitrogen are docuvely shown. The LOD of acetone (as ponditions is calculated. The ability to		

14. SUBJECT TERMS 15. NUMBER OF PAGES Methane Capillary waveguide Nd:YAG laser 12 Methane hydrates Fiber optic probe Toluene Raman spectroscopy 16. PRICE CODE Acetone 17. SECURITY CLASSIFICATION 18. SECURITY CLASSIFICATION 19. SECURITY CLASSIFICATION OF REPORT 20. LIMITATION OF ABSTRACT OF THIS PAGE OF ABSTRACT **UNCLASSIFIED** UNCLASSIFIED **UNCLASSIFIED** UL

detect all three states (gas, liquid, and solid) is of great importance to the detection and characterization of ocean floor methane.

CONTENTS

INTRODUCTION	1
EXPERIMENTAL	2
RESULTS	4
CONCLUSIONS	9
REFERENCES	9

FIBER OPTIC RAMAN SPECTROSCOPY FOR DETECTION OF METHANE HYDRATES AND RELATED SPECIES

INTRODUCTION

The detection of methane hydrates in the deep ocean environment is essential for the characterization of potential natural gas resources. Current technologies being deployed suffer from limitations including non-specificity, complexity, and slow response times. Optical methods afford the most flexibility with respect to methane hydrate detection. There are three regimes in which methane detection is desired: as a solid, as a gas in water, and dissolved in water. Optical spectroscopy allows measurements to be made in all three scenarios. In particular, Raman spectroscopy provides a wealth of analytical information regarding not only methane but many related hydrocarbons (ethane, propane, etc.) and potentially other species that contaminate the hydrates. Knowledge of the type and quality of hydrate present at a given location would be invaluable for the assessment of various locations of hydrated natural gas.

Initial studies to determine the feasibility of using Raman spectroscopy for the analysis of methane hydrates indicate that Raman probes may be useful in the characterization of oceanic environments including gaseous, liquid methane in sea water and methane hydrate. Raman spectroscopy is a well-established technique and is an attractive approach for deep oceanic environments due to its remote operability either through a window or when coupled to fiber optics. The use of optical fibers is ideal for ocean deployment for two reasons: 1) Measurements can be directed towards an area of interest using a robotic arm, and 2) damage can be restricted to the fiber optic probe not the laser and spectrometer. Due to the inherently weak nature of Raman scattering, a capillary waveguide approach is taken to increase the collected Raman scatter of the sample.

We are proposing to use the second harmonic (532 nm, 18,797 cm⁻¹) of a Nd:YAG laser for Raman excitation of methane C-H vibrational bands. The detection of other small hydrocarbons (ethane, propane, etc.) that often indicate the presence of a methane source will be investigated through excitation of C-C and -CH₃ vibrational modes. The use of a miniature spectrometer will be investigated and tested with respect to sensitivity and resolution. Initially, liquid organic materials have been studied neat and diluted in water as a simple substitute for methane, whose solubility is very low at atmospheric pressure, rendering the concentration too low to be measured with the available detector. A list of the normalized Raman cross sections (N_{σ}) is given in Table 1 for comparison.^{3,4} The Raman cross section is a measure of the efficiency of inelastic photon scattering versus the elastic Rayleigh photon scattering which produces no change in frequency. For example, when compared with nitrogen, methane has a Raman cross section which is 9.0 times greater, acetone has a cross section which is 2.4 times greater, and water's Raman cross section is 3.4 times greater. Due to its lower relative Raman cross section and methyl groups, and high solubility in water acetone was chosen initially as a substitute for methane.

Manuscript approved November 26, 2001.

Table 1. Relative Normalized Differential Raman Scattering Cross Sections (N_{\odot}) for Vibrational Bands of Various Molecules

Molecule	v _j (cm ⁻¹)	Exciting λ (nm)	Normalized σ	Ratio Nσ(X) / Nσ(CH₄)
N ₂	2331	515	1.0	0.1
H ₂	4156	515	3.4	0.4
GF _M	2017	56	47 - 19 0 - 14	PARTUO PART
C ₂ H ₆	993	488	1.2	0.1
C₃H ₈	2890	51 5	6.6	0.7
	867	5 15	1.7	0.2
H₂O	3652	515	3.4	0.4
CH ₃ OH	2955	515	10.9	1.2
	2846	515	6.9	8.0
C₂H₅OH	2943	515	25.6	2.8
CHCl₃	773	515	1.6	0.2
CCI ₄	459	515	6.95	8.0
C_6H_6	3070	515	15.3	1.7
	992	515	12.1	1.3
C_6H_{12}	802	515	4.1	0.5
(CH ₃) ₂ CO	782	488	2.4	0.3

EXPERIMENTAL

The system used for feasibility testing consisted of a Nd: Yag Laser operating at 532 nm (18,797 cm-1) with a power of 35 mW at a repetition rate of 15 Hz and a pulse width of 3-5 ns. Light was injected into a 600 µm excitation fiber (FVP600630660, Polymicro Technologies, Phoenix, AZ) using a 200 mm focal length lens (Model No. 41380, Oriel, Stamford, CT). The fiber was held in place using a fiber chuck (Newport, CA) mate to a X-Y positioner (New Focus, CA) to align the fiber face with the excitation beam. All incident excitation energies were measured at the distal end of the excitation fiber using a detector head (LM-1, Coherent, Auburn, CA) and a digital meter (Fieldmaster GS, Coherent, Auburn, CA). The 600 µm excitation fiber was one of two fibers in dual parallel fiber design probe, one delivering light and the other 600 µm collection fiber gathering the Raman scatter. The collection fiber was terminated using a SMA-905 fiber optic connector interfaced with a fiber optic breakout filter holder containing a holographic 532 nm notch filter. A 400 µm fiber on the opposite end of the filter holder delivered the Rayleigh line-filtered Raman scatter to an miniature spectrometer (PC2000, Ocean Optics, Dunedin, FL), an uncooled linear CCD array, via another SMA-905 fiber optic connector using an integration time of 5 sec. Spectrometer slits were not present, thus the input optical fiber (400 µm) defined the effective slit resulting in a resolution of approximately 150 cm⁻¹ (FWHM). While this represents low resolution Raman spectroscopy, identification of organic species is readily possible as will be demonstrated in the following section. A diagram of the optical layout of the system is given in Figure 1. All measurements were made using the capillary waveguide, which consisted of a 2 mm i.d., 2.4 mm o.d. fused silica tube (Vitrocom Inc, Mt. Lakes, NJ). The ends of the dual fiber optic probe were inserted into the end of the capillary waveguide as depicted in Figure 2.

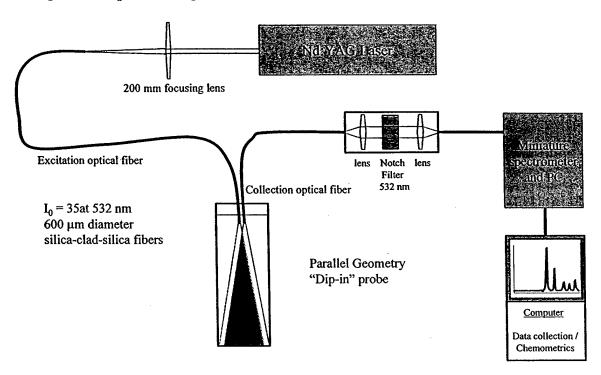


Figure 1. Nd:YAG laser using the 2nd harmonic output coupled to a fiber optic probe with a fiber breakout and notch filter for Raman spectroscopy

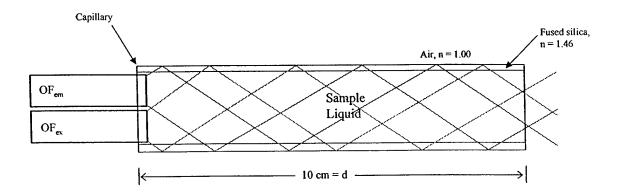


Figure 2. Capillary waveguide showing the rays of light emanating from the excitation fiber (OF_{ex}) and collection or emission fiber (OF_{em})

RESULTS

The use of a capillary waveguide / dual fiber probe combination in air with the sample solution inside the capillary enables a significant increase in signal to be realized at very little cost to the end user. This can be seen in Figure 3, where spectra of a neat solution of acetonitrile have been taken using a two fiber parallel "dip-in" probe with and without a capillary waveguide attached. The effect of the capillary is a four-fold increase in signal: for the 2935 cm⁻¹ band of acetonitrile, the two fiber probe signal level was 525 counts versus the signal obtained using the capillary waveguide, 2050 counts.

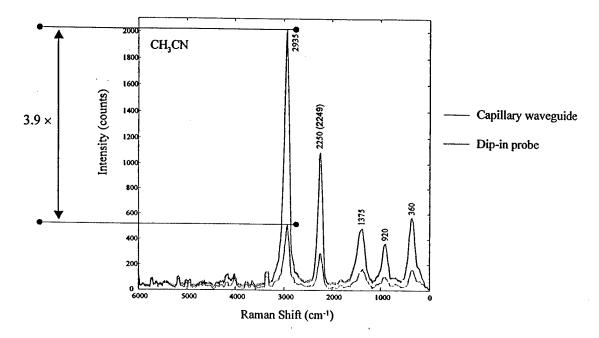


Figure 3. Capillary waveguide enhancement of Raman signal of an acetonitrile (CH₃CN) solution

The ability to measure low resolution Raman spectra using an inexpensive uncooled linear CCD can be seen in Figures 4 and 5. Many bands are visible in the Raman spectrum of neat toluene in Figure 4, as are visible in the spectrum of neat chloroform (CHCl₃) in Figure 5. The Raman spectrum of a mixture of hexane isomers is shown in Figure 6. Five bands in the spectrum of hexane are easily identifiable, even at this reduced resolution. The separately measured overlaid Raman spectra of water and acetone, in Figure 7, shows that they should easily be identifiable in a mixture.

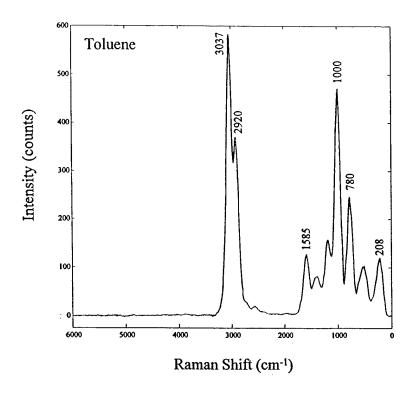


Figure 4. Raman spectrum of neat toluene using the capillary waveguide fiber optic system. Three spectra are measured in series are overlaid in the plot

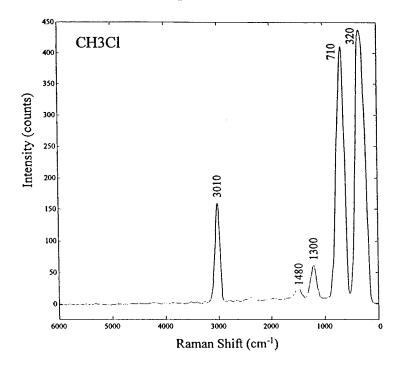


Figure 5. Raman spectrum of neat chloroform (CHCl₃) using the capillary waveguide fiber optic system

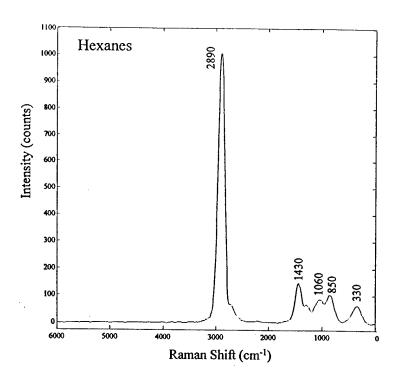


Figure 6. Raman spectrum of a mixture of neat hexane isomers using the capillary waveguide fiber optic system

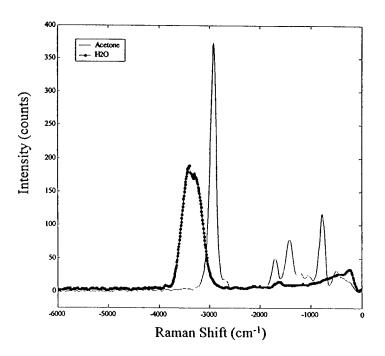


Figure 7. Overlaid Raman spectra of water (dots) and acetone (line) using the capillary waveguide fiber optic system

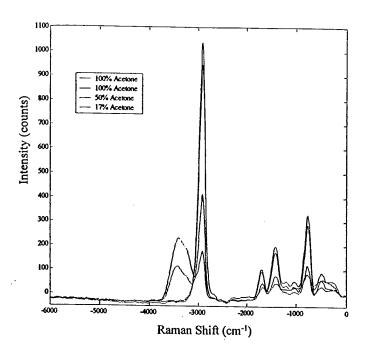


Figure 8. Raman spectra of acetone and water mixtures measured using the capillary waveguide fiber optic system

The spectra of several concentrations of acetone in water are given in Figure 8. Using acetone in water as a surrogate a calibration curve was generated and the resulting acetone calibration curve is shown in Figure 9. Accounting for the difference in the Raman cross-sections, the estimated limits of detection (LOD) and limits of quantitation (LOQ) for methane were calculated assuming higher laser powers and integration times. The use of a new detector which has better charge coupled device (CCD) sensitivity and a light collection lens provides a factor of 50 in terms of sensitivity according to manufacturers specifications. This improvement combined with integration time and laser power increases will permit the detection of methane in this application. These calculated LOD are consistent with the concentrations of methane expected at depth and are summarized in Table 2.

Table 2. Experimental Modifications and LOD Improvement of Acetone and Methane

	Improvement				
Experimental modification	Factor	LOD (%)	LOQ (%)		
Acetone with current detector, 35mW, 5 s integration tin	-	2.1	6.9		
Acetone with new detector, 35mW, 5 s integration time	50	4.2×10^{-2}	1.4×10^{-1}		
Acetone with new detector, 100mW, 5 s integration time	145	1.4×10^{-2}	4.7×10^{-1}		
Acetone with new detector, 100mW, 10 s integration tim	290	7.2×10^{-3}	2.3×10^{-2}		
Methane with new detector, 100mW, 5 s integration time		3.6×10^{-3}	1.2×10^{-1}		
Methane with new detector, 100mW, 10 s integration time		1.9×10^{-3}	6.2×10^{-2}		

^{*} LOD - limit of detection; LOD is baseline + 3s

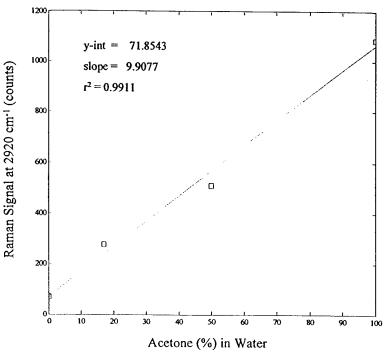


Figure 9. Analytical calibration curve for acetone in water

^{*} LOQ - Limit of quantitation; LOQ is baseline + 10s

CONCLUSIONS

The use of a capillary waveguide provides a significant increase in the measured Raman signal from organic molecules. This optical configuration combined with several improvements/modifications in the analysis method and instrumentation, will permit the measurement of methane and related molecules at depth.

REFERENCES

^{1.} S. Dai, J. P. Young, G. M. Begun, J. E. Coffield, and G. Mamantov, *Applied Spectroscopy*, 46, 375, 1992.

^{2.} Jie Lin, Sean J. Hart, Jonathan E. Kenny, "Improved Two Fiber Probe for in situ Spectroscopic Analysis", *Analytical Chemistry*, **1996**, 68, 3098-3103.

^{3.} A. Weber, <u>Raman Spectroscopy of Gases and Liquids</u>, Springer-Verlag, New York, 1979.

^{4.} D. P. Strommen, K. Nakamoto, <u>Laboratory Raman Spectroscopy</u>, John Wiley & Sons, New York, 1984.